CRC: Project Proposal

Jennifer Rathe, PGY-2 Pediatrics. August 22, 2013.

A. Study Purpose and Rationale

The purpose of this study is to determine the functionally conserved and variable regions of human rhinovirus (HRV) within the viral swarm infecting human subjects.

Human rhinovirus (HRV) is a single-stranded RNA virus, which causes upper respiratory infections and exacerbations of asthma and chronic obstructive pulmonary disease. It is the most frequent cause of the common cold and asthma exacerbations, costing billions every year in medications, hospital stays, and missed work and school days (1-8).

HRV has > 100 strain or lineages within 3 different species and design of vaccines and anti-viral medications largely unsuccessful (9, 10). With the numerous HRV strains with differing immunogenic capsids and host cell receptor tropism, targeting the virus is a challenge. However, even well-designed and targeted medications have largely failed to limit or stop infection. The failure of these past medications is likely due to an RNA viral evolutionary strategy called quasispecies (11-13).

Human rhinovirus (HRV) is theorized to infect human hosts as a quasispecies, which enables an evolutionary advantage. The main or 'consensus' virus mutates with every replication cycle generating a swarm of mutated, but related viral genomes. The host places pressures on the swarm, rather than on individual virus particles thus enabling functional advantages to a swarm of genetically different viral genomes vs. identical viral genome copies (14-17).

Over the last 5 years, methods have been established to investigate quasispecies within natural HRV infections. The protocols enable visualization of hundreds to thousands of copies of the viral swarm (18, 19). Using next generation sequencing, a small sample of 15 natural HRV infections have shown variability as well as conserved regions within the swarm of individual infections. However, this study has been limited by the small number of samples and that all the viruses were of the same strain of HRV.

My project will analyze ~200 HRV samples for conserved and variable regions across swarms representing multiple HRV lineages. I hypothesize that despite numerous and genetically variable lineages, that the swarms will reveal conserved regions for functional experiments and ultimately, better targets for anti-viral development.

B. Sample Ascertainment and Characteristics

- 1. Collected from groups around the world with URI/respiratory symptoms of varying severity and no restrictions on race, age, gender, or medical problems. [Requirements were simply HRV viral load high enough to be considered infectious and nasal swabs or lavage that had not been cultured.]
- 2. Samples were sequenced via 454 and Illumina next generation sequencing by Institute for Genome Sciences at the University of Maryland School of Medicine.
- 3. The sequences have been assembled into consensus sequences and are publicly available on NCBI databases.

C. Study Design and Statistical Analysis

- 1. Swarm variant detection with gsVariant and illumina software
- Alignment of 'master' swarm variant file against relative consensus HRV strain [hrv39 swarm against HRV 39 consensus...] [Programs → HMMER and Se-Al]
- 3. Location of all swarm variants [insertions, deletions, recombinations, and SNPs] along a master genome alignment [Programs → excel and R]

- 4. Distribution of all variants across 200 viral infection samples, determination of 99th and 1st percentile values [Prism]
- 5. Apply 99th and 1st percentile values to graph created in Step 3 above, and identify regions that are hyper-variable and conserved [greater than 99% and less than 1%]
- 6. Assess these regions for function associated with conservation or hypervariability [RNA folding, protein on capsid, polymerase, etc.]
- 7. Test region(s) of interest in functional in vitro experiments [mutate virus un region of interest and compare viral replication/survival/swarm mutations to control virus]

D. Limitations

- 1. Samples are already sequenced and many only cover 80-90% of genome [may not have swarm data for entire HRV genome]
- 2. The 99th and 1st percentile cut-offs may be too stringent or too liberal to have a reasonable number of regions to investigate [if this happens, then the cut off can be adjusted and/or regions with a greater number of conserved variants screened]
- 3. More sequences may be needed to define areas of conservation, may just be a level of noise covering a large portion of genome

- 1. Bardin, P.G., Fraenkel, D.J., Sanderson, G., Dorward, M., Lau, L.C., Johnston, S.L., and Holgate, S.T. 1994. Amplified rhinovirus colds in atopic subjects. *Clin Exp Allergy* 24:457-464.
- 2. Dreschers, S., Dumitru, C.A., Adams, C., and Gulbins, E. 2007. The cold case: are rhinoviruses perfectly adapted pathogens? *Cell Mol Life Sci* 64:181-191.
- Johnston, S.L., Pattemore, P.K., Sanderson, G., Smith, S., Lampe, F., Josephs, L., Symington, P., O'Toole, S., Myint, S.H., Tyrrell, D.A., et al. 1995. Community study of role of viral infections in exacerbations of asthma in 9-11 year old children. *Bmj* 310:1225-1229.
- 4. Khetsuriani, N., Lu, X., Teague, W.G., Kazerouni, N., Anderson, L.J., and Erdman, D.D. 2008. Novel human rhinoviruses and exacerbation of asthma in children. *Emerg Infect Dis* 14:1793-1796.
- 5. Kistler, A., Avila, P.C., Rouskin, S., Wang, D., Ward, T., Yagi, S., Schnurr, D., Ganem, D., DeRisi, J.L., and Boushey, H.A. 2007. Pan-viral screening of respiratory tract infections in adults with and without asthma reveals unexpected human coronavirus and human rhinovirus diversity. *J Infect Dis* 196:817-825.
- Lau, S.K., Yip, C.C., Tsoi, H.W., Lee, R.A., So, L.Y., Lau, Y.L., Chan, K.H., Woo, P.C., and Yuen, K.Y. 2007. Clinical features and complete genome characterization of a distinct human rhinovirus (HRV) genetic cluster, probably representing a previously undetected HRV species, HRV-C, associated with acute respiratory illness in children. J Clin Microbiol 45:3655-3664.

- Miller, E.K., Edwards, K.M., Weinberg, G.A., Iwane, M.K., Griffin, M.R., Hall, C.B., Zhu, Y., Szilagyi, P.G., Morin, L.L., Heil, L.H., et al. 2008. A novel group of rhinoviruses is associated with asthma hospitalizations. J Allergy Clin Immunol.
- 8. Camargo, C.A., Jr., Ginde, A.A., Clark, S., Cartwright, C.P., Falsey, A.R., and Niewoehner, D.E. 2008. Viral pathogens in acute exacerbations of chronic obstructive pulmonary disease. Intern Emerg Med 3:355-359.
- Palmenberg, A.C., Spiro, D., Kuzmickas, R., Wang, S., Djikeng, A., Rathe, J.A., Fraser-Liggett, C.M., and Liggett, S.B. 2009. Sequencing and analyses of all known human rhinovirus genomes reveal structure and evolution. Science 324:55-59.
- Hayden, F.G., Turner, R.B., Gwaltney, J.M., Chi-Burris, K., Gersten, M., Hsyu, P., Patick, A.K., Smith, G.J., 3rd, and Zalman, L.S. 2003. Phase II, randomized, double-blind, placebo-controlled studies of ruprintrivir nasal spray 2-percent suspension for prevention and treatment of experimentally induced rhinovirus colds in healthy volunteers. Antimicrob Agents Chemother 47:3907-3916.
- 11. Florea, N.R., Maglio, D., and Nicolau, D.P. 2003. Pleconaril, a novel antipicornaviral agent. *Pharmacotherapy* 23:339-348.
- Hayden, F.G., Herrington, D.T., Coats, T.L., Kim, K., Cooper, E.C., Villano, S.A., Liu, S., Hudson, S., Pevear, D.C., Collett, M., et al. 2003. Efficacy and safety of oral pleconaril for treatment of colds due to picornaviruses in adults: results of 2 double-blind, randomized, placebo-controlled trials. *Clin Infect Dis* 36:1523-1532.

- 13. Ledford, R.M., Patel, N.R., Demenczuk, T.M., Watanyar, A., Herbertz, T., Collett, M.S., and Pevear, D.C. 2004. VP1 sequencing of all human rhinovirus serotypes: insights into genus phylogeny and susceptibility to antiviral capsid-binding compounds. J Virol 78:3663-3674.
- 14. Bull, J.J., Meyers, L.A., and Lachmann, M. 2005. Quasispecies made simple. *PLoS Comput Biol* 1:e61.
- 15. Eigen, M. 1993. Viral quasispecies. Sci Am 269:42-49.
- 16. Holland, J.J., De La Torre, J.C., and Steinhauer, D.A. 1992. RNA virus populations as quasispecies. *Curr Top Microbiol Immunol* 176:1-20.
- 17. Vignuzzi, M., Stone, J.K., Arnold, J.J., Cameron, C.E., and Andino, R. 2006. Quasispecies diversity determines pathogenesis through cooperative interactions in a viral population. *Nature* 439:344-348.
- Cordey, S., Junier, T., Gerlach, D., Gobbini, F., Farinelli, L., Zdobnov, E.M., Winther, B., Tapparel, C., and Kaiser, L. 2010. Rhinovirus genome evolution during experimental human infection. *PLoS ONE* 5:e10588.
- 19. Wright, C.F., Morelli, M.J., Thebaud, G., Knowles, N.J., Herzyk, P., Paton, D.J., Haydon, D.T., and King, D.P. 2010. Beyond the consensus: dissecting within-host viral population diversity of foot-and-mouth disease virus by using next-generation genome sequencing. J Virol 85:2266-2275.